



AGRI MAGAZINE

(International E-Magazine for Agricultural Articles)

Volume: 03, Issue: 05 (May, 2026)

Available online at <http://www.agrimagazine.in>

© Agri Magazine, ISSN: 3048-8656

Polyploidy Breeding

Dr. Sunidhi Tiwari¹, *Arya Subandhu², A. Srivani², C.V Abhinetra Reddy², Chirag T.S²,
V.Shabarish² and M.R Aseesh Gupta²

¹Assistant Professor, Faculty of Agriculture, Jagannath University, Jaipur, India

²Student, B.Sc. (Hons.) Agriculture, Jagannath University, Jaipur, India

*Corresponding Author's email: aryasubandhu03@gmail.com

Polyploidy breeding is recognized as a fundamental tool in crop improvement, involving the multiplication of complete sets of chromosomes [1]. The evolution of approximately 70% of angiosperms is attributed to natural polyploidization events [2]. Higher genetic diversity and phenotypic plasticity are provided by polyploidy, enabling plants to adapt to diverse ecological niches [3]. The manipulation of ploidy levels is utilized by researchers to create novel germplasm with enhanced economic value [4].

Classification and Biological Basis

Autopolyploidy

Autopolyploidy is originated from the duplication of a single genome belonging to the same species [5].

The 'gigas' effect, characterized by an increase in cell volume and organ size, is frequently observed in autopolyploid plants [3].

Seedless varieties of fruits, such as triploid watermelons, are successfully developed through autopolyploidy breeding [6].

The formation of multivalents during meiosis is identified as a primary cause of reduced fertility in these lines [5].

Allopolyploidy

Allopolyploidy is resulted from the hybridization of two or more distinct species followed by chromosome doubling [7].

Meiotic stability is often achieved in allopolyploids due to the presence of non-homologous genomes which favor bivalent pairing [8].

Significant crops such as bread wheat (hexaploid), upland cotton (tetraploid), and canola are classified as allopolyploids [2].

The combination of divergent gene pools within a single nucleus is facilitated by allopolyploidy, leading to heterosis [7].

Induction Methods and Agronomic Applications

Chemical Induction Techniques

Colchicine is extensively employed as a spindle inhibitor to induce chromosome doubling in actively dividing tissues [9].

Specific concentrations of colchicine are applied to seeds, axillary buds, or callus cultures to generate polyploid sectors [10].

Alternative anti-microtubule agents, including oryzalin and trifluralin, are increasingly used due to their lower toxicity [11].

Impact on Crop Quality and Resistance

Increased concentrations of essential nutrients and secondary metabolites are reported in polyploid medicinal plants [12].

Enhanced resistance to abiotic stresses, particularly drought and salinity, is frequently exhibited by polyploid genotypes [13].

The vegetative growth phase is often extended in polyploids, which is beneficial for biomass production in forage crops [1].

Suggestion

The integration of flow cytometry for rapid ploidy determination is suggested to enhance selection efficiency in large breeding populations [14].

More focus should be directed towards the induction of polyploidy in underutilized legumes to improve their yield potential [13].

The use of tissue culture-based induction methods is proposed to minimize the occurrence of chimeric plants [10].

Polyploidy breeding should be combined with CRISPR/Cas9 technology to precisely target gene duplications for crop improvement [4].

References

1. Singh, B. D. (2022). *Plant Breeding: Principles and Methods*. Kalyani Publishers.
2. Wendel, J. F. (2000). *Genome evolution in polyploids*. Plant Molecular Biology.
3. Stebbins, G. L. (1950). *Variation and Evolution in Plants*. Columbia University Press.
4. Comai, L. (2005). The advantages and disadvantages of being polyploid. *Nature Reviews Genetics*.
5. Ramsey, J., & Schemske, D. W. (1998). Pathways and mechanisms of polyploid formation.
6. Kihara, H. (1951). Triploid watermelons. *Horticultural Science*.
7. Feldman, M., & Levy, A. A. (2012). Genome evolution in wheat allopolyploids.
8. Udall, J. A., & Wendel, J. F. (2006). Polyploidy and plant breeding. *Crop Science*.
9. Blakeslee, A. F., & Avery, A. G. (1937). Methods of inducing doubling of chromosomes.
10. Dhooche, E. et al. (2011). Mitotic blocking agents for in vitro polyploidy induction.
11. Kermani, M. J. et al. (2003). Oryzalin as an alternative to colchicine in ornamental plants.
12. Iannicelli, J. et al. (2020). Polyploidy to improve medicinal plants. *Frontiers in Plant Science*.
13. Dhawan, O. P., & Lavania, U. C. (1996). Enhancing productivity via induced polyploidy.
14. Dolezel, J. et al. (2007). Plant DNA flow cytometry and estimation of genome size.