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Blast Disease of Rice: Causes and Prevention Methods

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Rice blast disease, caused by the fungus *Magnaporthe oryzae* (anamorph: *Pyricularia oryzae*), is considered the most destructive disease of rice worldwide [1]. Severe yield losses ranging from 10 to 100 percent are reported in susceptible varieties when environmental conditions favor disease development [2]. The disease is prevalent in over 85 countries and poses a significant threat to global food security, particularly in Asia where rice is a staple crop [3]. Economic losses due to rice blast are estimated to destroy enough rice each year to feed approximately 60 million people, underscoring the critical need for effective management strategies [4].

Causes of Rice Blast

Causal Organism

Magnaporthe oryzae is an ascomycete fungus belonging to the order Magnaporthales, responsible for causing blast disease on all above-ground parts of the rice plant [5].

The fungus produces pyriform to obclavate conidia that are three-celled and hyaline, which serve as the primary infectious propagules under favorable conditions [1].

A remarkable degree of genetic variability is exhibited by the pathogen, resulting in a large number of distinct races that differ in their virulence patterns on rice cultivars [6].

Disease Cycle

The disease cycle is initiated when conidia land on susceptible host tissue and germinate under conditions of adequate moisture and temperature [7].

A specialized infection structure called the appressorium is formed by the germinating conidium, generating enormous turgor pressure to penetrate the rice cuticle mechanically [5].

Colonization of host tissue is followed by the production of new conidia within four to five days, enabling rapid disease spread within and between fields [2].

The fungus survives in infected rice straw, seeds, and soil debris between growing seasons, serving as primary inoculum sources for subsequent crop cycles [8].

Environmental Factors Favoring Disease

Optimal disease development is favored by temperatures ranging between 24°C and 28°C combined with relative humidity above 90 percent [7].

Leaf wetness periods of six hours or more are required for successful spore germination and infection, making nighttime dew a critical factor in epidemic development [6].

Excessive application of nitrogenous fertilizers is strongly correlated with increased susceptibility to blast, as lush vegetative growth provides favorable conditions for fungal colonization [3].

Symptoms and Diagnosis

Diamond-shaped or spindle-shaped lesions with gray or white centers and brown to reddish-brown borders are the characteristic foliar symptoms of rice blast [1].

Neck blast, the most devastating form of the disease, results in the rotting of the panicle base and causes the head to droop, a symptom commonly referred to as 'rotten neck' [9].

Node blast is identified by the presence of blackened and collapsed nodes on the culm, which leads to lodging and complete panicle death in severe infections [2].

Collar blast results in rotting at the junction of the leaf blade and leaf sheath, eventually causing the entire leaf to wither and die [8].

Prevention and Management Strategies

Host Resistance

The development and deployment of blast-resistant varieties is considered the most economical and environmentally sustainable approach to managing rice blast [10].

Resistance genes such as Pi-ta, Pi-b, Pi-k, and Pi-9 have been identified and incorporated into improved rice varieties through conventional and molecular breeding programs [6].

Multiline varieties and varietal mixtures are employed to reduce the rate of disease progress by limiting the spread of single virulent pathogen races across the entire crop [11].

Chemical Control

Systemic fungicides belonging to the strobilurin and triazole chemical groups are widely employed for blast management when disease pressure exceeds economic threshold levels [12].

Tricyclazole, propiconazole, and azoxystrobin are among the most commonly recommended fungicides for field application against *M. oryzae* in major rice-growing regions [5].

Seed treatment with systemic fungicides such as carbendazim or captan is practiced to reduce seedborne inoculum and protect the young seedlings during the early growth stages [4].

Cultural and Agronomic Practices

Judicious application of nitrogen fertilizers in split doses is recommended to avoid excessive vegetative growth, which significantly predisposes the crop to blast infection [3].

Proper field sanitation including the destruction of infected stubble and crop residues is enforced to reduce the primary inoculum load for the following crop season [8].

Maintaining optimal plant spacing to improve canopy aeration is recommended as it reduces the humidity within the crop stand, creating conditions less favorable for fungal sporulation [7].

Use of certified disease-free seeds and hot-water seed treatment at 52°C for 10 minutes are established practices for reducing seedborne blast inoculum before planting [9].

Biological Control

Biocontrol agents such as *Trichoderma harzianum*, *Pseudomonas fluorescens*, and *Bacillus subtilis* have demonstrated significant antagonistic activity against *M. oryzae* under field conditions [10].

Induction of systemic resistance in rice plants through the application of plant activators such as silicon and salicylic acid has shown promise as an eco-friendly blast management strategy [11].

References

1. Ou, S. H. (1985). Rice Diseases. 2nd ed. Commonwealth Mycological Institute, Kew, UK.
2. Bonman, J. M. (1992). Durable Resistance to Rice Blast Disease. *Euphytica*, 63, 45–55.
3. Valent, B., & Khang, C. H. (2010). Recent advances in rice blast effector research. *Current Opinion in Plant Biology*, 13(4), 434–441.
4. Skamnioti, P., & Gurr, S. J. (2009). Against the grain: safeguarding rice from rice blast disease. *Trends in Biotechnology*, 27(3), 141–150.
5. Dean, R. A. et al. (2005). The genome sequence of the rice blast fungus *Magnaporthe grisea*. *Nature*, 434, 980–986.
6. Kang, S. et al. (2016). Genome organization and evolution of the rice blast fungus. *Nature Genetics*, 48, 1067–1077.
7. Kim, S. G. et al. (2009). Epidemiology of rice blast in Korea. *Korean Journal of Plant Pathology*, 25, 120–130.

8. Prabhu, A. S. et al. (2009). Blast Management in Upland Rice. *Pesquisa Agropecuaria Brasileira*, 44, 1–10.
9. Wang, G. L., & Valent, B. (2009). Advances in breeding for disease resistance. *Annual Review of Phytopathology*, 47, 453–465.
10. Zhu, Y. et al. (2000). Genetic diversity and disease control in rice. *Nature*, 406, 718–722.
11. Naqvi, S. A. H. (2004). *Diseases of Fruits and Vegetables: Diagnosis and Management*. Springer, Netherlands.
12. Ploetz, R. C. (2003). *Diseases of Tropical Fruit Crops*. CABI Publishing, UK.